Assay: VURP5_BR
(Viral Upper Respiratory Panel 5 for BioRad)

Quantity: 100 x 20µL PCR reactions

5-plex assay: SARS-CoV-2, Influenza A, Influenza B, Respiratory syncytial virus A and B, and human RPP30 DNA

SKU: PNP-VURP5-D-BR-100 (BioRad)

(RUO). Research Use Only. Not for use in Diagnostic Procedures.

SCOPE OF THIS DOCUMENT:

The oligonucleotide recipes are optimized for each instrument (Bio-Rad, QuantStudio). The verification data presented in this product information sheet were obtained using BUN-VURP5-D-BR-100 on a Bio-Rad CFX96. The performance of the other SKUs on their corresponding instrument should be similar. Contact PCRassays.com if you need to use a different qPCR instrument.

CONTENTS

The primers and probes in the VURP5 assay are provided in Tube 1 as a 5X concentrated working solution that detects 4 pathogens and a human extraction control.

Table of Dyes used in this assay:

Pathogen/Target	Dyes	Quencher	Refs.
Flu A	FAM	BHQ-1	1,2
RPP30-DNA control	HEX	BHQ-1	7
SARS-CoV-2	TEX615	BHQ-2	3,4
RSV (A and B)	Cy5	BHQ-2	5
Flu B	Cy5.5	BHQ-2	6

The probes are designed as TaqMan[8] cleavage mechanism and thus the reaction requires a DNA polymerase with 5'-exonuclease activity.

ASSAY HANDLING AND CONTAMINATION

The VURP5 bundle is shipped on ice, and should be stored at -20 °C. The tubes should be kept on ice once thawed. Do not subject the enzyme to multiple freeze-thaw cycles.

Any contamination should be avoided by using appropriate personal protective equipment (PPE), powder free gloves, aerosol barrier pipette tips, and a clean hood.

Assay contents:

Tube 1: Primer/Probe mix (5X) for SARS-CoV-2, Flu A, Flu B, RSV A/B, and human RPP30 DNA.

Tube 2: (**Do NOT add to specimen unknowns**) Positive control: Synthetic 500 bp DNA fragments of Flu A, SARS-CoV-2, RSVA/B, Flu B, and human RPP30 DNA.

Tube 3: InhibiTaq RT-qPCR enzyme Mastermix (enough for 100 rxns. with 20 µL total volume). This is a custom formulation from Fortis Life Sciences to the specifications of PCRassays.com.

EXPERIMENTAL

Perform nucleic acid extraction/purification (recommended).

Set up your reaction (20 µL) as follows on ice:

Component	Volume (µL)
InhibiTaq RT-qPCR enzyme mastermix (2X)	10
Primer/Probe mix (5X)	4
Sample	2
Water (Molecular Biology Grade)	4

Notes: To improve assay sensitivity, up to 6 µL of sample can be added (water volume adjusted accordingly) for a total reaction volume of 20 µL.
For positive control rxns., add 2 µL of the solution from Tube 2.
For 10 µL reactions, divide all of the amounts above by a factor of 2.

A PCR protocol was used for verification on a Bio-Rad CFX96, with the following program:

Step	Thermocycling Protocol:
1	Incubate @ 50°C for 10 minutes
2	Incubate @ 95°C for 3 minutes
3	Incubate @ 95°C for 5 seconds
4	Incubate @ 55°C for 15 seconds
5	Plate Read
6	Go to step 3, repeat 44x more

RESULT INTERPRETATION

After running the qPCR reaction, use the instrument software to determine the quantification cycle, C_q (or use C_T if your instrument does not have the capability to compute a C_q). Fluorescence channels with a C_q < 38 cycles, and final RFU >Threshold are considered "positive" or "+" in the Table below. The "Threshold" value for calling a PCR positive is dependent on the instrument model, well size, and sample volume; thus the user must determine the threshold that is appropriate for their method. For our Bio-Rad CFX-96 instrument with 100 µL wells and 20 µL reaction volume, the average RFU was approximately 3,000 and we used a threshold of 300 for calling positives or "+" in the Table below.

Flu A FAM ™	SARS-CoV-2 TEX 615™	RSV A/B Cy5 ™	Flu B Cy5.5 ™	RPP30 HEX™	Recommended Interpretation
–	–	–	-	–	The PCR reaction failed. Please repeat the experiment.
–	–	–	-	+	The sample contains human RPP30 DNA. The sample doesn't contain viral RNA.
+	–	–	-	–	The sample contains Flu A RNA. The sample may not contain human RPP30 DNA.
+	–	–	-	+	The sample contains Flu A RNA and human RPP30 DNA.
–	+	–	-	–	The sample contains SARS-CoV-2 RNA. The sample may not contain human RPP30 DNA.
–	+	–	-	+	The sample contains SARS-CoV-2 RNA and human RPP30 DNA.
–	–	+	-	–	The sample contains RSV RNA. The sample may not contain human RPP30 DNA.
–	–	+	-	+	The sample contains RSV RNA and human RPP30 DNA.
-	-	-	+	-	The sample contains Flu B RNA. The sample may not contain human RPP30 DNA.
-	-	-	+	+	The sample contains Flu B RNA and human RPP30 DNA.
+	+	+	+	–	The sample contains Flu A RNA, SARS-CoV-2 RNA, RSV RNA, and Flu B RNA.The sample may not contain human RPP30 DNA.
+	+	+	+	+	The sample contains Flu A RNA, SARS-CoV-2 RNA, RSV RNA, Flu B RNA, and human RPP30 DNA.

VERIFICATION EXPERIMENTS

The VURP5 assay verification was carried out as a 5-plex assay, which simultaneously detects RNA from Influenza A, SARS-CoV-2, Respiratory syncytial virus, Influenza B, and human RPP30 DNA, which serves as a positive control assay.

Experiments were performed in triplicate using the experimental procedure given above, but with different samples added to each reaction. The samples used for the verification experiments contained 1×10^4 copies/reaction of synthetic 500 bp DNA constructs (from Twist Biosciences) harboring the regions of interest from the pathogen genomes: human RPP30 DNA gene, full-length synthetic SARS CoV2 RNA (Twist – control 50, Omicron variant), quantitative genomic RNA obtained from ATCC: FluA H1N1 (VR-1894DQ), FluA H3N2 (VR-1882DQ), RSVA (VR-1540DQ), RSVB (VR-1803D), Flu B (VR-1804DQ). **Figure 1** shows the results of these experiments, which indicate that the 5-plex specifically detects the different pathogens.

The limit of detection (LOD) was estimated by performing serial dilution experiments in triplicate (**Figure 2**). For dilution series only target construct was added. The results show a limit of detection (LOD) <10 copies/reaction.

Conclusion: The data in **Figure 1** indicate that the 5-plex primers and probes specifically detect and differentiate the pathogens and are also compatible with RPP30_DNA positive control primers. Human genomic DNA doesn't interfere with the detection of the pathogens.

NOTES

[1] FAM™ (Carboxyfluorescein), a trademark of Life Tech. Corporation.

[2] BHQ-1™ (Black Hole Quencher) a trademark of Biosearch Tech., Inc.

[3] TEX615™ is a trademark of Thermo Fisher Scientific.

[4] BHQ-2™ (Black Hole Quencher) a trademark of Biosearch Tech., Inc.

[5] Cy5™, a trademark of GE Healthcare.

[6] Cy5.5, a trademark of GE Healthcare.

[7] HEX™ (Hexachloro-fluorescein) a trademark of ThermoFisher Sci.

[8] TaqMan™ is a trademark of Roche Diagnostics, Inc.

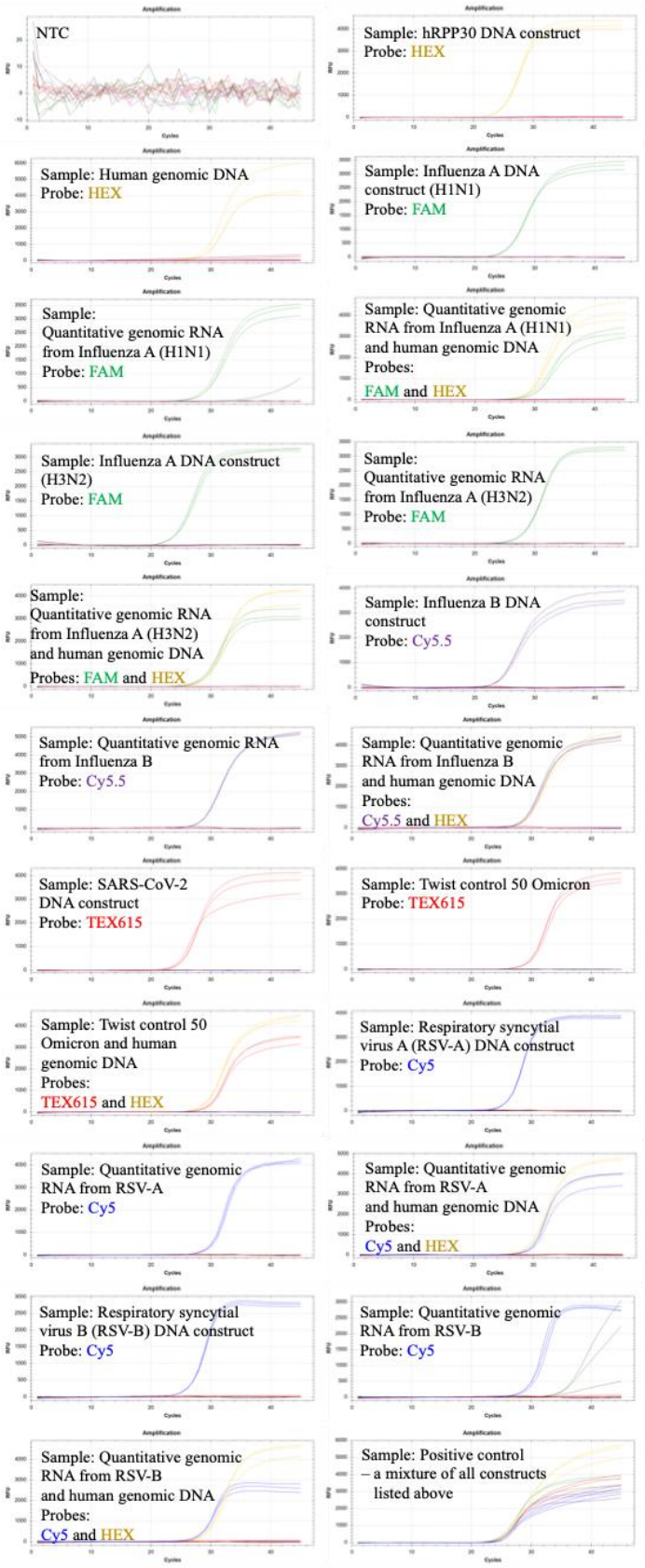

Figure 1: Verification experiments with single and multiple targets (given in text boxes for each panel). All sets of probes and primers are present in every reaction, but positive signal is only observed when the target(s) is present, indicating that the amplification is specific.

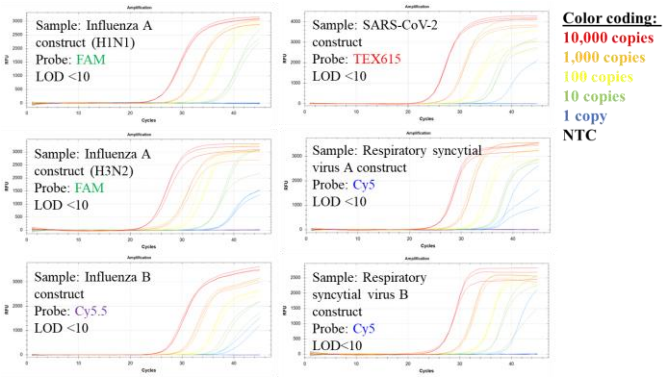

Figure 2: Serial dilution experiments show LOD <10 molecules for the synthetic DNA construct of each target.

CONTACT US

For assistance, please contact DNA Software using the link: https://www.pcrassays.com/contact/

Address: Michigan Life Science and Innovation Center,
46701 Commerce Center Dr, Plymouth, MI 48170

Phone: (734) 222-9080